

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
7 August 2003 (07.08.2003)

PCT

(10) International Publication Number
WO 03/063847 A1

(51) International Patent Classification⁷: **A61K 31/05**,
31/192, 35/78, A61P 1/08

(21) International Application Number: PCT/GB03/00451

(22) International Filing Date: 3 February 2003 (03.02.2003)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
0202385.1 1 February 2002 (01.02.2002) GB
0206183.6 15 March 2002 (15.03.2002) GB

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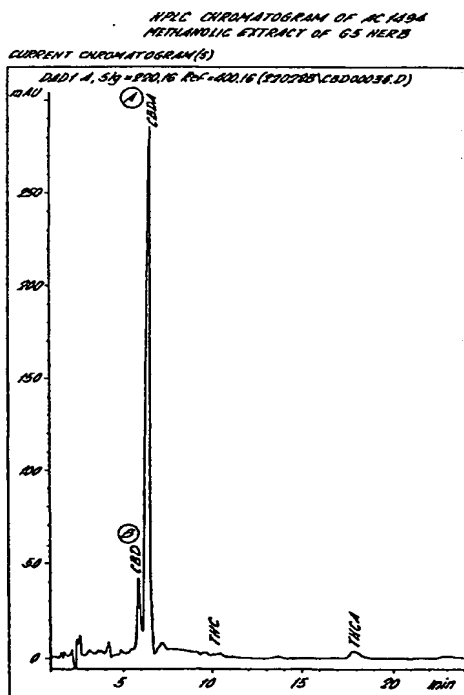
(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW),

[Continued on next page]

(54) Title: COMPOSITIONS COMPRISING CANNABINOIDS FOR TREATMENT OF NAUSEA, VOMITING, EMESIS, MOTION SICKNESS OR LIKE CONDITIONS

(57) Abstract: Cannabinoids, in particular CBD and CBDA and their acid derivatives are provided for use as an active pharmaceutical substance in the treatment of nausea, vomiting, emesis, motion sickness. In particular extracts of cannabis plants are presented which are rich in these substances and suitable for pharmaceutical use.



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Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM),
European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE,
ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, SE, SI,
SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN,
GQ, GW, ML, MR, NE, SN, TD, TG).

Declarations under Rule 4.17:

- as to applicant's entitlement to apply for and be granted a patent (Rule 4.17(ii)) for the following designations AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, UZ, VC, VN, YU, ZA, ZM, ZW, ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent

(AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG)

- of inventorship (Rule 4.17(iv)) for US only

Published:

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

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COMPOSITIONS COMPRISING CANNABINOIDS FOR TREATMENT OF NAUSEA, VOMITING, EMESIS,
MOTION SICKNESS OR LIKE CONDITIONS

5 Functional vomiting is the forceful expulsion of
gastric contents produced by involuntary contraction
of the abdominal musculature. This occurs when the
gastric fundus and lower oesophageal sphincter are
relaxed. Functional vomiting may be accompanied by
10 nausea (an unpleasant feeling that vomiting is about
to occur). Nausea is associated with altered
physiological activity, including gastric
hypomotility, and increased parasympathetic tone.
Nausea may precede and accompany vomiting. They
15 represent the patient's awareness of afferent stimuli
to the medullary vomiting centre.

Physiological vomiting is a functional condition that
occurs in response to a number of factors affecting
20 the vomiting centre. It may also be triggered by
peripheral factors such as ingestion of toxins,
disturbance of the vestibular system, peritoneal
inflammation and bowel obstruction. It may also occur
in disorders of delayed gastric emptying as, for
25 example, in diabetes and idiopathic gastroparesis.

Psychogenic vomiting may be self-induced or may occur
involuntarily in situations that are anxiety inducing,
threatening or in some way regarded as distasteful by
30 the subject. It is also possible that psychological
factors leading to vomiting are culturally determined
(for example, eating exotic food may be considered
repulsive in the subject's own cultural group).
Vomiting may also express hostility as when children
35 vomit during a temper tantrum or in certain conversion
disorders.

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Nausea and vomiting may also be induced by cytotoxic chemotherapy and radiotherapy. Post-operatively, patients may also vomit and experience nausea, which may be attributable to the anaesthetic and analgesic agents frequently administered concurrently.

There are therefore peripheral and central mechanisms which are involved in the expression of nausea and frank vomiting. Existing therapies are available for the treatment of these conditions but they have limitations, and there is a need for alternative treatments, particularly where these can exert their effect through a central nervous mechanism.

Investigation of a number of agents in a conscious animal model in which motion sickness is induced has confirmed that extracts of cannabis have an anti-emetic effect. Conventionally, the anti-emetic effect of cannabis has been ascribed to delta⁹-tetrahydrocannabinol (Δ^9 -THC). The use of a whole animal, conscious model to explore this effect, and the availability of cannabis extracts containing predominantly one cannabinoid or another have allowed for more detailed analysis of the contribution of specific cannabinoids.

Surprisingly, it has been found that the anti-emetic effect, in a model of motion sickness in *Suncus murinus* (the Asian musk (house) shrew), is greatest in high cannabidiol (CBD) (and / or its acid form CBDA) containing extracts rather than in high tetrahydrocannabinol (THC) (and / or its acid form THCA) containing extracts (greater than 50% CBD more preferably greater than 80% most preferably greater than 90% relative to other cannabinoids present).

It is hypothesised that the results will extend to the

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propyl variant of CBD, namely CBDV and its acid form CBDVA.

5 More particularly, the results were noted in extracts in which the cannabinoids were predominantly in their acid form since the extracts were prepared by a methanolic extraction and had not been subjected to a decarboxylation step by, for example, heating. It is possible therefore that the therapeutic effects noted are due to the acid form of the cannabinoids. If it is 10 the acid form of the cannabinoid that is responsible for the observed therapeutic effect this is particularly surprising since the acid forms of cannabinoids have not hitherto been known to exhibit therapeutic effects.

15 According to a first aspect of the present invention there is provided a cannabis extract, rich in CBD and / or CBDA and / or the propyl variants CBDV and / or CBDVA, for use in the manufacture of a medicament for 20 the treatment of nausea, vomiting, emesis, motion sickness or like conditions.

By rich is meant greater than 2% w/w of CBD and / or CBDA and / or the propyl variants CBDV and / or CBDVA, 25 more particularly greater than 5%, more preferably still greater than 7%.

30 According to another aspect of the present invention there is provided the use of CBD and / or CBDV in the manufacture of a medicament for the treatment of nausea, vomiting, emesis, motion sickness or like conditions.

35 According to another aspect of the present invention there is provided a cannabinoid acid for use as an active pharmaceutical substance.

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According to yet another aspect of the present invention there is provided CBDA or CBDVA for use as an active pharmaceutical substance.

- 5 Preferably the active pharmaceutical substance is present as a medicament for the treatment of nausea, vomiting, emesis, motion sickness or like conditions.

10 In one embodiment the CBD and / or CBDA and / or CBDV and / or CBDVA are present with other cannabinoids as a mixture derived from a plant extract (CMBE, cannabis based medicinal extract).

15 Plant extracts are preferred as, in addition to one or more cannabinoids, they will contain other chemical entities that may provide a beneficial effect either in their own right or in combination with the one or more cannabinoids. Such other chemicals include, for example, volatile oils e.g. terpene or carotene rich
20 volatiles. Known terpenes present in the CMBE include C_{10} terpenes, e.g. mycerene, and pinenes and C_{15} terpenes e.g. caryophyllene.

25 Preferably the CBD or CBDV and / or the acids thereof are present with THC or THCV and / or the acids thereof.

30 Alternatively the CBD or CBDV and / or the acids thereof are substantially free (less than 10%, more preferably less than 5% and most preferably less than 2% relative to other cannabinoids present) from other cannabinoids.

35 In another embodiment the CBD or CBDV and / or the acids thereof are synthetic.

The invention also extends to methods of treating

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nausea, vomiting, emesis, motion sickness or like conditions with CBD or CBDV and / or the acids thereof, either as the sole active ingredient or in mixtures as plant extracts.

5

Whilst the observation has been made on an extract administered intraperitoneally, the skilled man will appreciate that a medicament can be prepared for administration by any suitable means. These include, but are not limited to, solids, semi solids, e.g. gels, liquids, sprays, aerosols, inhalers, vapourisers, enemas, rectal suppositories and the like. The route of administration need not be intraperitoneally but could be oral, buccal, sublingual, or by any other suitable route e.g. the respiratory tract, nasal tract and distal rectum.

A "plant extract" is an extract from a plant material as defined in the Guidance for Industry Botanical Drug Products Draft Guidance, August 2000, US Department of Health and Human Services, Food and Drug Administration Centre for Drug Evaluation and Research.

Plant material" is defined as a plant or plant part (e.g. bark, wood, leaves, stems, roots, flowers, fruits, seeds, berries or parts thereof) as well as exudates.

The term "Cannabis plant(s)" encompasses wild type Cannabis sativa and also variants thereof, including cannabis chemovars which naturally contain different amounts of the individual cannabinoids, Cannabis sativa subspecies indica including the variants var. indica and var. kafiristanica, Cannabis indica and also plants which are the result of genetic crosses, self-crosses or hybrids thereof. The term

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"Cannabis plant material" is to be interpreted accordingly as encompassing plant material derived from one or more cannabis plants. For the avoidance of doubt it is hereby stated that "cannabis plant material" includes dried cannabis biomass.

In the context of this application the terms "cannabis extract" or "extract from a cannabis plant", which are used interchangeably encompass "Botanical Drug Substances (BDS)" derived from cannabis plant material. A Botanical Drug Substance is defined in the Guidance for Industry Botanical Drug Products Draft Guidance, August 2000, US Department of Health and Human Services, Food and Drug Administration Centre for Drug Evaluation and Research as: "A drug substance derived from one or more plants, algae, or macroscopic fungi. It is prepared from botanical raw materials by one or more of the following processes: pulverisation, decoction, expression, aqueous extraction, ethanolic extraction, or other similar processes." A botanical drug substance does not include a highly purified or chemically modified substance derived from natural sources. Thus, in the case of cannabis, "botanical drug substances" derived from cannabis plants do not include highly purified, Pharmacopoeial grade cannabinoids.

"Botanical drug substances" derived from cannabis plants include primary extracts prepared by such processes as, for example, maceration, percolation, extraction with solvents such as C1 to C5 alcohols (e.g. ethanol), Norflurane (HFA134a), HFA227 and liquid carbon dioxide under pressure. The primary extract may be further purified for example by supercritical or subcritical extraction, vaporisation and chromatography. When solvents such as those listed above are used, the resultant extract contains

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non-specific lipid-soluble material. This can be removed by a variety of processes including "winterisation", which involves chilling to -20°C followed by filtration to remove waxy ballast, extraction with liquid carbon dioxide and by distillation.

Preferred "cannabis extracts" include those which are obtainable by using any of the methods or processes specifically disclosed herein for preparing extracts from cannabis plant material. The extracts are preferably substantially free of waxes and other non-specific lipid soluble material but preferably contain substantially all of the cannabinoids naturally present in the plant, most preferably in substantially the same ratios in which they occur in the intact cannabis plant.

Botanical drug substances are formulated into "Botanical Drug Products" which are defined in the Guidance for Industry Botanical Drug Products Draft Guidance, August 2000, US Department of Health and Human Services, Food and Drug Administration Centre for Drug Evaluation and Research as: "A botanical product that is intended for use as a drug; a drug product that is prepared from a botanical drug substance."

"Cannabinoids" may be highly purified, Pharmacopoeial Grade substances and may be obtained by purification from a natural source or via synthetic means. The cannabinoids will include, but are not limited to, tetrahydrocannabinoids, their precursors, alkyl (particularly propyl) analogues, cannabidiols, their precursors, alkyl (particularly propyl) analogues, and cannabiniol.

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In preferred embodiments of the invention the formulations comprise extracts of one or more varieties of whole Cannabis plants, particularly Cannabis sativa, Cannabis indica or plants which are the result of genetic crosses, self-crosses or hybrids thereof. The precise cannabinoid content of any particular cannabis variety may be qualitatively and quantitatively determined using methods well known to those skilled in the art, such as TLC or HPLC. Thus, one may chose a Cannabis variety from which to prepare an extract which will produce the desired ratio of CBD or CBDV to THC or THCV. Alternatively, extracts from two of more different varieties may be mixed or blended to produce a material with the preferred cannabinoid ratio for formulating into a pharmaceutical formulation.

The preparation of convenient ratios of CBD, CBDV, CBDA and CBDVA -containing medicines is made possible by the cultivation of specific chemovars of cannabis. These chemovars (plants distinguished by the cannabinoids produced, rather than the morphological characteristics of the plant) can be been bred by a variety of plant breeding techniques which will be familiar to a person skilled in the art. Suitable methods are given in Example 3. Propagation of the plants by cuttings for production material ensures that the genotype is fixed and that each crop of plants contains the cannabinoids in substantially the same ratio.

Horticulturally, it is convenient to grow chemovars producing e.g. CBD and CBDV as the predominant cannabinoid from cuttings. This ensures that the genotype in each crop is identical and the qualitative formulation (the proportion of each cannabinoid in the biomass) is the same. From these chemovars, extracts

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can be prepared by the similar method of extraction. Convenient methods of preparing primary extracts include maceration, percolation, extraction with solvents such as C1 to C5 alcohols (ethanol),

5 Norflurane (HFA134a), HFA227 and liquid carbon dioxide under pressure. The primary extract may be further purified for example by supercritical or subcritical extraction, vaporisation and chromatography. When solvents such as those listed above are used, the

10 resultant extract contains non-specific lipid-soluble material. This can be removed by a variety of processes including chilling to -20°C followed by filtration to remove waxy ballast, extraction with liquid carbon dioxide and by distillation. Preferred

15 plant cultivation and extract preparation methods are shown in the Examples. The resulting extract is suitable for incorporation into pharmaceutical preparations.

20 A detailed examination of the pharmacological differences between CBD and THC has revealed significant differences in these compounds and consequently the finding that CBD and / or its acid CBDA appear to be responsible for the therapeutic

25 effects noted was surprising. THC is bound with high avidity to CB1 and CB2 receptors in cerebral cortex and other sites; CBD is relatively inactive against CB1 receptors and appears to have non-cannabinoid receptor pharmacological actions in the central

30 nervous system. Without prejudice to the teaching of the invention, it is possible that the anti-emetic effect of CBD and / or its acid CBDA is mediated via a non-cannabinergic mechanism.

35 Table 1 below illustrates some of the differences between these cannabinoids.

Table 1:

	Effect	THC	THCV	CBD	CBDV	Reference
5	CB ₁ (Brain receptors) CB ₂ (Peripheral receptors)	++ +		± -		Pertwee et al, 1998
10	CNS Effects Anticonvulsant † Antimetrazol Anti-electroshock Muscle Relaxant	-- - - --		++ - ++ ++		Carlini et al, 1973 GW Data GW data Petro, 1980
15	Antinociceptive Catalepsy Psychoactive Antipsychotic Neuroprotective antioxidant activity*	++ ++ ++ - +		++ ++ - ++ ++		GW data GW data GW data Zuardi et al, 1991 Hampson A J et al, 1998
20	Antiemetic Sedation (reduced spontaneous activity) Appetite stimulation Appetite suppression	++ + ++		- + ++		Zuardi et al, 1991
25	Anxiolytic	-		++		GW data
30	Cardiovascular Effects Bradycardia Tachycardia Hypertension § Hypotension § Anti-inflammatory	- + + - ±		+ - - + ±		Smiley et al, 1976 Adams et al, 1977 Brown, 1998

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Table 1 Cont.

Immunomodulatory/anti-inflammatory activity					
Raw Paw Oedema Test	-		++		GW data
Cox 1					GW data
Cox 2					GW data
TNF α Antagonism	+	+	++	++	
Glaucoma	++		+		

* Effect is CB1 receptor independent.

† THC is pro convulsant

§ THC has a biphasic effect on blood pressure; in naïve patients it may produce postural hypotension and it has also been reported to produce hypertension on prolonged usage.

GW Internal Report No 002/000159.

Whilst it is known that THC can be used to control nausea and vomiting pre-operatively the effect of other cannabinoids or combinations or the effect of the acid forms present in, for example, plant extracts was not hitherto known.

The applicants studied the effect of other cannabinoids as cannabis extracts, and particularly extracts containing predominantly CBD or its acid form CBDA in *Suncus murinus* and in which an emetic response can be induced by a motion stimulus. Compounds which are effective in this test have therapeutic benefit in the treatment of motion-induced nausea and vomiting, and also these conditions when induced by other pathways in the peripheral and central nervous systems.

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The applicant has determined that, for example, extracts in which the content of CBD and / or CBDA is 2-20% w/w, and the content of THC and or THCA is 0.1 - 2% w/w are particularly beneficial.

5

The invention is further illustrated with reference to the accompanying figures and examples:

10 Fig 1 is a TLC from the methanolic extract of the high CBD chemovar G5 (M16). It shows significant CBD and CBDA peaks at around 6 and 7 minutes and lesser amounts of THC and THCA at around 10 and 18 minutes.

15 Fig 2 is the TLC the methanolic extract of the high THC chemovar G2 (M6). It shows significant THC and THCA peaks at around 10 and 18 minutes along with lesser amounts of CBD and CBDA at around 6 and 7 minutes.

20 The figures shown in Figs 1 and 2 are quantative.

Fig 3 is a thin layer chromatography plate showing the methanolic extracts, and by way of comparison BDS (decarboxylated and extracted by sub critical liquid CO₂) The results confirm that the methanolic extracts
25 comprise a high proportion of the respective cannabinoids THC and CBD in what was later shown to be their acid forms.

30 Fig 4 illustrates the results obtained for G5, the upper trace being for the BDS (decarboxylated and extracted by sub critical liquid CO₂) and the lower trace for the methanolic extract. The CBD peak at 35 minutes and the THC peak at 38 minutes are marked.

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Fig 5 illustrates the results obtained for G2, the upper trace being for the BDS (decarboxylated and extracted by sub critical liquid CO₂) and the lower trace for the methanolic extract. The CBD peak at 35 minutes and the THC peak at 37 minutes are marked.

Example 1:

Extracts of cannabis can be prepared by a number of solvent extraction techniques, including the use of organic solvents alone or in admixture with water and under sub critical or supercritical conditions. Cannabinoids may be present in cannabis biomass as free cannabinoids and as the corresponding acidic precursors. Conventional methods of preparation involve the total extraction of free cannabinoids and precursors with solvents such as lower alkyl alcohols, particularly methanol. In this example, total extracts of a high THC, and a high CBD-containing chemovar were made using methanol.

Biomass from each chemovar was separately extracted in a column with methanol at room temperature, and the pooled percolate was collected. Solvent was removed by evaporation in a rotary evaporator at a temperature not exceeding 43°C. The resulting high THC and high CBD extracts were dispersed in 5% Polysorbate 80/normal saline and a Polysorbate/saline vehicle was used as control. The high THC extract (M6) contained more than 10% of THC and / or THCA and less than 1% of CBD and / or CBDA. The high CBD extract (M16) contained more than 7.3% of CBD and / or CBDA and less than 2% of THC and / or THCA.

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A preliminary HPLC analysis of the methanolic extracts (and comparative data for a botanical drug substance (BDS), when prepared by sub critical CO₂ extraction after decarboxylation) is given in the table 2 below:

5

Table 2.

Extract/ Analyte	Methanolic extract from a CBD rich chemovar % w/w	Comparative Decarboxylated CO ₂ extract from THC rich chemovar % w/w	Comparative Decarboxylated CO ₂ extract from CBD rich chemovar % w/w
THC	N.D.	64.2%	2.9%
THCA	1.4%	N.D.	N.D.
CBD	6.1%	1.1%	70.2%
CBDA	49.9%	N.D.	N.D.
CBN	N.D.	1.0%	N.D.

10

As would be expected, the acid forms of the cannabinoid predominate in the methanolic extract of the non-decarboxylated herb.

15

For the methanolic extracts the % w/w of the principal cannabinoid (sum of acid + neutral forms) is lower than found in the equivalent BDS. Again, this is expected due to the lower selectivity of the methanol, as compared to liquid CO₂. Thus, the methanolic extraction gives rise to the extraction of more non-target material and a diluting of the active content of the extract.

20

25

The content of the methanolic extract was confirmed by TLC analysis and chromatogram traces for the high THC and high CBD extracts are shown in Figs 1 and 2.

30

- 15 -

The traces in Figs 1 and 2 indicate the acid form of the cannabinoid to be the principal component observed, with smaller amounts of the corresponding neutral cannabinoid also being detected.

5

Samples of the methanolic extracts were run on a TLC plate along with some cannabinoid markers, and by way of comparison botanic drug substance (BDS) (decarboxylated and extracted by sub critical liquid CO₂). The results are illustrated in Fig 3.

10

Further analysis of the methanolic extracts, and by way of comparison the BDS (decarboxylated and extracted by sub critical liquid CO₂), using gas chromatography indicates the presence of terpenes at around the 14 to 18 minute mark. It should be noted that because G.C is operated at around 250°C any acid form of the CBD and THC present in the extracts will be decarboxylated and appear on the trace as CBD or THC. The results are illustrated in Figs 4 and 5.

15

20

The most significant difference between the lower traces of Fig 4 and 5 is the difference between the THC and CBD peaks.

25

Example 2:

30

Adult *Suncus murinus* (30-89 g bodyweight) of either sex were injected with either G2 (M6) or G5 (M16) at a range of doses from 1.0, 2.0 or 4.0 mg/kg (or vehicle), intraperitoneally, 30 minutes prior to a motion stimulus (1 Hz 40 mm amplitude of shaking for 10 minutes). The animals were observed for any overt behavioural change. A number of emetic episodes and

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the latency of onset were recorded. Data were expressed as the mean \pm s.e.m., group size $n = 6$ and statistically analysed using ANOVA, followed by Bonferroni Dunnett's T test. The results are summarised in table 3 below.

Table 3: Effect of Cannabis Extracts given Intraperitoneally on Frequency of Latency to First Emesis (means \pm s.e.m.)

Treatment	Dose mg/kg	Emetic Episodes, mean \pm s.e.m.			
		Latency (secs)	P value	Number	P Value
Vehicle control		103.0 \pm 25.3		12.4 \pm 2.4	
M16	1.0	206.8 \pm 33.9	<0.05	6.3 \pm 1.4	<0.05
High CBD chemovar	2.0	250.5 \pm 84.5	<0.05	6.6 \pm 2.4	<0.05
	4.0	Increase	N5	Increase	N5
M6	1.0	† NDDC		NDDC	
High THC chemovar	2.0	NDDC		NDDC	
	4.0*	<100		>18	<0.05

† NDDC no detectable difference from control

* Increase in vomiting, reduction in latency

The results for the high CBD producing chemovar extract (M16) shown in Table 3 are not unlike U-shaped dose response curves noted for some other pharmacological actions of cannabinoids.

The lack of effect of the high THC producing chemovar extract in this test system is surprising. Indeed, at high doses there appears to be an increase in vomiting and reduction in latency. It was confirmed that neither M6 nor M16 had emetic activity in their own right. These data further emphasise the differences noted in the pharmacological effects of THC and CBD,

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which have been investigated by the applicant.

Example 3 Growing of Medicinal Cannabis

Plants are grown as clones from germinated seeds,
under glass at a temperature of $25^{\circ}\text{C} \pm 1.5^{\circ}\text{C}$ for 3
weeks in 24-hour daylight; this keeps the plants in a
vegetative state. Flowering is induced by exposure to
12 hour day length for 8-9 weeks.

10 No artificial pesticides, herbicides, insecticides or fumigants are used. Plants are grown organically, with biological control of insect pests.

The essential steps in production from seed accession
15 to dried Medicinal Cannabis are summarised as follows:

```

Seed Accessions
|
20 Seeds germinated at G-Pharm (UK)
|
Selection for cannabinoid content and vigour
|
Mother Plant
|
25 Cuttings rooted
14-21 days in peat plug
25 C, 24 hour day length
|
30 Rooted cuttings potted up in 5 litre pots of bespoke
compost
|
Young Clone Plant established
3 weeks, 24 hour day length, 25 °C
|
35 Lower Branches Removed end of week 3
Used to make new generation of cuttings
|

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Induction of flowering

Plant relocation to 12 hour day length are to induce flowering

↓

Flower formation and maturation

8-9 weeks at 25 °C

↓

Harvest

90% of flowers and leaves senesced

↓

Drying

Under conditions of light exclusion

↓

MEDICINAL CANNABIS

15

**Example 4 Determination of Cannabinoid Content in
Plants and Extracts**

Identity by TLC

20

a) Materials and methods

Equipment Application device capable of delivering an
accurately controlled volume of solution i.e

25 1 µl capillary pipette or micro litre syringe.

TLC development tank with lid

Hot air blower

30

Silica gel G TLC plates (SIL N-HR/UV254),
200 µm layer with fluorescent indicator on
polyester support.

35

Dipping tank for visualisation reagent.

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Mobile phase 80% petroleum ether 60:80/20% Diethyl ether.

5 Visualisation reagent 0.1% w/v aqueous Fast Blue B
(100mg in 100ml de-ionised water). An optional method is to scan at UV 254 and 365 nm.

10 b) Sample preparation

i) Herbal raw material

Approximately 200mg of finely ground, dried cannabis is weighed into a 10ml volumetric flask. Make up to
15 volume using methanol:chloroform (9:1) extraction solvent.

Extract by ultrasound for 15 minutes. Decant supernatant and use directly for chromatography.

20

ii) Herbal drug Extract

Approximately 50mg of extract is weighed into a 25ml volumetric flask. Make up to volume using
25 methanol solvent. Shake vigorously to dissolve and then use directly for chromatography.

c) Standards

30 0.1 mg/ml delta-9-THC in methanol.
0.1mg/ml CBD in methanol.

The standard solutions are stored frozen at -20°C between uses and are used for up to 12 months after

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initial preparation.

d) Test solutions and method

5 Apply to points separated by a minimum of 10mm.

- 10 i) either 5 μ l of herb extract or 1 μ l of
 herbal extract solution as appropriate,
 ii) 10 μ l of 0.1 mg/ml delta-9-THC in methanol
 standard solution,
 iii) 10 μ l of 0.1mg/ml CBD in methanol standard
 solution.

15 Elute the TLC plate through a distance of 8cm,
 then remove the plate. Allow solvent to
 evaporate from the plate and then repeat the
 elution for a second time (double development).

20 The plate is briefly immersed in the Fast Blue B
 reagent until the characteristic re/orange colour
 of cannabinoids begins to develop. The plate is
 removed and allowed to dry under ambient
 conditions in the dark.

25 A permanent record of the result is made either
 by reproduction of the image by digital
 scanner(preferred option) or by noting spot
 positions and colours on a tracing paper.

30 Assay THC, THCA, CBD, CBDA and CBN by HPLC

a) Materials and methods

Equipment: HP 1100 HPLC with diode array detector
 and autosampler. The equipment is set

- 21 -

up and operated in accordance with
in-house standard operating procedures
(SOPlab037)

5	HPLC column	Discovery C8 5 μ m, 15x 0.46 cm plus Kingsorb ODS2 precolumn 5 μ m 3 x 0.46 cm.
10	Mobile Phase	Acetonitrile:methanol:0.25% aqueous acetic acid (16:7:6 by volume)
	Column Operating Temperature	25°C
	Flow Rate	1.0 ml/min
15	Injection Volume	10 μ l
	Run time	25mins
20	Detection	Neutral and acid cannabinoids 220nm (band width 16nm) Reference wavelength 400nm/bandwidth 16nm
25		Slit 4nm
30		Acid cannabinoids are routinely monitored at 310nm (band width 16nm) for qualitative confirmatory and identification purposes only.
	Data capture	HP Chemistation with Version A7.01 software

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b) Sample preparation

5 Approximately 40mg of Cannabis Based
Medicinal Extract is dissolved in 25ml
methanol and this solution is diluted
to 1 to 10 in methanol. This dilution
is used for chromatography.

10 0.5 ml of the fill solution, contained
within the Pump Action Sublingual Spray
unit, is sampled by glass pipette. The
solution is diluted into a 25ml flask
and made to the mark with methanol.
15 200 μ l of this solution is diluted with
800 μ l of methanol.

20 Herb or resin samples are prepared by
taking a 100mg sample and treating this
with 5 or 10ml of Methanol/Chloroform
(9/1 w/v). The dispersion is sonicated
in a sealed tube for 10 minutes,
allowed to cool and an aliquot is
centrifuged and suitably diluted with
methanol prior to chromatography.

25

c) Standards

30 External standardisation is used for this method.
Dilution of stock standards of THC, CBD and CBN in
methanol or ethanol are made to give final working
standards of approximately accurately 0.1 mg/ml. The
working standards are stored at -20°C and are used for
up to 12 months after initial preparation.

- 23 -

Injection of each standard is made in triplicate prior to the injection of any test solution. At suitable intervals during the processing of test solutions, repeat injections of standards are made. In the
5 absence of reliable CBDA and THCA standards, these compounds are analysed using respectively the CBD and THC standard response factors.

The elution order has been determined as CBD, CBDA,
10 CBN, THC and THCA. Other cannabinoids are detected using this method and may be identified and determined as necessary.

d) Test solutions
15

Diluted test solutions are made up in methanol and should contain analytes in the linear working range of 0.02-0.2 mg/ml.

20 e) Chromatography Acceptance Criteria:

The following acceptance criteria are applied to the results of each sequence as they have been found to result in adequate resolution of all analytes
25 (including the two most closely eluting analytes CBD and CBDA)

i) Retention time windows for each analyte:
30 CBD 5.4-5.9 minutes
 CBN 7.9-8.7 minutes
 THC 9.6-10.6 minutes

ii) Peak shape (symmetry factor according to BP

- 24 -

method)

CBD < 1.30

CBN < 1.25

5 THC < 1.35

10 iii) A number of modifications to the standard method have been developed to deal with those samples which contain late eluting impurity peaks e.g method CBD2A extends the run time to 50 minutes. All solutions should be clarified by centrifugation before being transferred into autosampler vials sealed with teflon faced septum seal and
15 cap.

20 iv) The precolumn is critical to the quality of the chromatography and should be changed when the back pressure rises above 71 bar and/or acceptance criteria regarding retention time and resolution, fall outside their specified limits.

25 f) Data Processing

Cannabinoids can be subdivided into neutral and acidic- the qualitative identification can be performed using the DAD dual wavelength mode. Acidic cannabinoids absorb strongly in the region of
30 220nm-310nm. Neutral cannabinoids only absorb strongly in the region of 220nm.

Routinely, only the data recorded at 220 nm is used for quantitative analysis.

- 25 -

The DAD can also be set up to take UV spectral scans of each peak, which can then be stored in a spectral library and used for identification purposes.

- 5 Data processing for quantitation utilises batch processing software on the Hewlett Packard Chemstation.

a) Sample Chromatograms

- 10 HPLC sample chromatograms for THC and CBD Herbal Drug extracts are provided in the accompanying Figures.

Example 5 Preparation of the Herbal Drug Extract

15

A flow chart showing the process of manufacture of extract from the High-THC and High-CBD (non acid and acid forms) chemovars is given below:

- 20 Medicinal Cannabis (High-THC or High-CBD)

↓

Chopping to predominantly 2 to 3mm

↓

Optional decarboxylation step:

- 25 Heating at 100 to 150°C for sufficient time to decarboxylate acid form of cannabinoids to produce neutral cannabinoids

↓

- 30 Extraction with a specified volume of liquid carbon dioxide over 6 to 8 hours

↓

Removal of CO₂ by depressurisation to recover crude extract

- 26 -

↓

"Winterisation"-Dissolution of crude extract in ethanol Ph. Eur. followed by chilling solution (-20°C/48 hrs) to precipitate unwanted waxes

5

↓

Removal of unwanted waxy material by cold filtration

↓

Removal of ethanol from the filtrate by thin film evaporation under reduced pressure

10

Example 6

High CBD cannabis was grown under glass at a mean temperature of 21 + 2°C, RH 50-60%. Herb was harvested and dried at ambient room temperature at a RH of 40-45% in the dark. When dry, the leaf and flower head were stripped from stem and this dried biomass is referred to as "medicinal cannabis".

Medicinal cannabis was reduced to a coarse powder (particles passing through a 3 mm mesh) and packed into the chamber of a Supercritical Fluid Extractor. Packing density was 0.3 and liquid carbon dioxide at a pressure of 600 bar was passed through the mass at a temperature of 35°C. Supercritical extraction is carried out for 4 hours and the extract was recovered by stepwise decompression into a collection vessel. The resulting green-brown oily resinous extract is further purified. When dissolved in ethanol BP (2 parts) and subjected to a temperature of -20°C for 24 hours a deposit (consisting of fat-soluble, waxy material) was thrown out of solution and was removed by filtration. Solvent was removed at low pressure in

- 27 -

a rotary evaporator. The resulting extract is a soft extract which contains approximately 60% CBD with up to 4% tetrahydrocannabinol, within a total of other cannabinoids of 6%. Extracts were made using THCV and
5 CBDV chemovars using the general method described above.

A high THC chemovar was similarly treated and yielded an extract containing approximately 60% THC and
10 approximately 6% of other cannabinoids of which 1-2 % is cannabidiol and the remainder is minor cannabinoids including cannabinol. Quantitative yield was 9% w/w based on weight of dry medicinal cannabis.

15 A person skilled in the art will appreciate that other combinations of temperature and pressure (in the range +10°C to 35°C and 60 - 600 bar) can be used to prepare extracts under supercritical and subcritical conditions.

20

Example 7-Preparation of CBDA

Summary of process:

25 Extract CBD herb (G5 chemovar) with 0.1% v/v acetic acid in hexane.

↓

Filter and remove solvent from filtrate on rotary evaporator.

30

↓

Pass a solution of the resulting CBDA enriched extract through a column packed with Sephadex LH20, eluting with 2:1 chloroform/dichloromethane.

↓

- 28 -

Collect CBDA rich fractions and remove solvent by rotary evaporation.

↓

5 Re-dissolve crude CBDA in methanol and remove insoluble residue by filtration.

↓

Remove solvent from filtrate by rotary evaporation.

↓

10 Re-dissolve crude CBDA in pentane and remove insoluble residue by filtration.

↓

Remove solvent from filtrate by rotary evaporation.

↓

CBDA crystals

15

Yield:

100 g of G5 chemovar yields approx 5 g of purified CBDA.

20

Characteristics:

Pale yellow crystalline solid

Melting Point = 45-48°C

25

Chromatographic purity = 94 % CBDA by area normalisation

*CBD 3 %.

30 THCA non detected i.e. < 0.1 %

THC non detected i.e. < 0.1 %

Material slowly decarboxylates in solution

CBDA → CBD + CO₂

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* As CBDA does not co-elute with CBD during processing of the extract in the low pressure column chromatography method employed, the detected CBD is likely to be formed from the breakdown of the CBDA during processing and analysis. This undesirable decarboxylation of the purified material might be minimised by manipulation of CBDA at sub-ambient temperatures.

10 **Example 8-Purification of CBD**

Overview of process

Starting from freshly harvested plant material the process comprises drying and decarboxylation of the plant material, optional treatment (e.g. milling) of the dried plant material to reduce the particle size (preferably to less than 2000µm), extraction with liquid carbon dioxide, ethanolic precipitation to reduce the amount of non-target material, clean-up of the crude ethanolic extract by passage through activated charcoal, removal of solvent (ethanol) to produce a CBD-enriched fraction, and re-crystallisation of CBD from pentane.

25 Plant material

GW Pharma Ltd has developed distinct varieties of Cannabis plant hybrids to maximise the output of the specific chemical constituents, cannabinoids. A "high CBD" chemovar designated G5 produces >90% total cannabinoid content as CBD (naturally occurring in the plant in the form of CBDA). Alternative "high CBD" varieties can be obtained - see for example, Common cannabinoids phenotypes in 350 stocks of cannabis, Small and Beckstead, Lloydia vol 36b , 1973 p144-156 -

- 30 -

and bred using techniques well known to the skilled man to maximise cannabinoid content.

Solvents

5 All solvents used in the isolation and analysis of CBD (e.g n-pentane) were, unless otherwise stated, of chromatographic or A.R. grade.

Standards

10 Reference materials from Sigma were used as standards in the analysis of extracts, intermediates and finished products, these were: Δ^9 THC in methanol BN 10601/B (ca. 1 mg/ml) and CBD in methanol BN 10601/C (ca. 1 mg/ml).

15

Preparation of a cannabidiol-containing extract

A cannabidiol-containing extract is prepared from a "high CBD" cannabis chemovar according to the following process:

20

Prepare ethanolic solution of botanical drug substance as follows:

harvest cannabis plant material, dry, reduce particle
25 size by milling to less than 2000 μ m

↓

decarboxylate milled plant material by heating to
approximately 105°C for 15 minutes, followed by
approximately 145°C for minimum of 55 minutes (NB
30 decarboxylation time and temperature may be varied)

↓

extract with liquid carbon dioxide (CO₂) [Food Grade]
for up to 10 hours Conditions: Approximately 60 bar \pm
10 bar pressure and 10°C \pm 5°C

35

↓

- 31 -

Removal of CO₂ by depressurisation to recover crude
extract

↓

5 "Winterisation"-Dissolution of crude extract in
ethanol followed by chilling solution
(-20°C ± 5°C/up to 52hours) to precipitate unwanted
waxes

↓

10 Removal of unwanted waxy material by cold filtration
(20mm filter)

↓

ethanolic solution of BDS
(Stored at -20°C ± 5°C)

15 Extraction using liquid CO₂ is carried out under
sub-critical conditions at a temperature of
approximately 10°C ± 5°C using a pressure of
approximately 60 bar ± 10 bar. Decarboxylated plant
20 material is packed into a single column and exposed to
liquid CO₂ under pressure for approximately 8 hours,
CO₂ mass flow 1250 kg/hr ± 20%.

Following depressurisation and venting off of the
CO₂ the crude BDS extract is collected into sealed
25 vessels. The crude BDS extract is held at -20°C ±
5°C.

The crude BDS extract contains waxes and long
chain molecules. Removal is by "winterisation",
30 whereby the crude BDS extract is warmed to e.g. 40°C ±
4°C to liquefy the material. Ethanol is added in the
ratio of 2:1 ethanol volume to weight of crude BDS
extract. The ethanolic solution is then cooled to
-20°C ± 5°C and held at this temperature for
35 approximately 48 hours.

- 32 -

On completion of the winterisation the precipitate is removed by cold filtration through a 20µm filter, to give an ethanolic solution of the BDS.

5 Preliminary charcoal clean-up may be carried out by passing the ethanolic BDS solution (400-500 mg/ml) through a disposable plastic column (130 mm x 27 mm i.d) packed with activated charcoal (decolourcarb DCL GDC grade, from Sutcliffe Speakman Carbons, 15.4 g per
10 unit). Absolute ethanol B.P. (Hayman) is used as the solvent.

Ethanol and any water that may be present are removed by rotary evaporation or thin film evaporation
15 under reduced pressure (60°C ± 2°C, with vapour at 40°C ± 2°C / 172 mbar and 72 mbar ± 4mbar) to produce a CBD-rich extract.

Solvent re-crystallisation

20 The CBD-rich extract is re-dissolving in a suitable solvent (e.g. n-pentane) and filtered to remove insoluble material. Solvent is then removed, e.g. by rotary evaporation, to produce crystalline CBD. All steps are carried out according to standard
25 laboratory procedures, such as would be known to those skilled in the art.

Product characteristics

Yield:

30 3 g of CBD BDS yields approx 1 g of purified CBD.

Characteristics:

White crystalline solid.

35 Chromatographic purity > 99% CBD by area normalization.

- 33 -

Chromatographic purity superior to commercially available CBD Sigma standard (refer to Figures 1 and 3).

- 5 THC non detected i.e. < 0.1%
 CBN non detected i.e. < 0.1%

Identity confirmed by HPLC, GC and TLC retention behaviour compared to CBD Sigma standard.

10

Assay vs both Sigma CBD std in range 98.0-102.0%

Melting Point = 64-66°C (literature value = 66-67°C).

15

HPLC analysis

The composition of the isolated products may be determined by HPLC analysis.

- 20 A typical HPLC assay for Δ^9 THC, Δ^9 THCA, CBD, CBDA and CBN may be carried out as follows:

a) Materials and methods

- 25 Chromatography Equipment and conditions:

Equipment Agilent (HP)1100 HPLC system with
 variable wavelength UV detector or
 diode array detector.

HPLC Column Discovery C8 5 μ m 15cm x 0.46cm

- 30 Pre-Column Kingsorb C18 5 μ m 3cm x 0.46cm

Mobile Phase Acetonitrile : Methanol : 0.25% w/v
 acetic acid (16:7:6 by volume)

Column Temp 25°C

Flow Rate 1.0ml min⁻¹

- 35 Detection 220nm 600mA f.s.d. Second wavelength
 310nm

Injection Volume 10 μ l

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Run Time 20-25 minutes (may be extended for
samples containing small amount of
late-eluting peaks)
Elution Order CBD, CBDA, Δ^9 THCV, CBN, Δ^9 THC, CBC, Δ^9
5 THCA

b) Sample preparation

10 Samples of "pure" cannabidiol are diluted in methanol
prior to HPLC analysis. Optimal dilutions may be
determined empirically.

15 Herbal cannabis samples are prepared by taking a 100mg
sample and treating this with 5 or 10ml of
Methanol/Chloroform (9/1 w/v). The dispersion is
sonicated in a sealed tube for 10 minutes, allowed to
cool and an aliquot is centrifuged and suitably
diluted with methanol prior to chromatography.

20

c) Standards

25 Stock standard solutions of CBD, CBN and Δ^9 THC in
methanol at approximately 1mg ml⁻¹ are stored at
-20°C. Diluted working standards (0.1 mg/ml for Δ^9
THC and CBD and 0.01 mg/ml for CBN) are prepared in
methanol from the stock standards and stored at -20°C
(maximum period of twelve months after initial
preparation). After preparation, standard solutions
30 must be aliquoted into vials to reduce the amount of
standard exposed to room temperature. Prior to use in
an HPLC sample assay, the required number of standard
vials are removed and allowed to equilibrate to room
temperature.

35

Injection of each standard is made in triplicate prior
to the injection of any test solution. At suitable
intervals during the processing of test solutions,

- 35 -

repeat injections of standards are made. In the absence of reliable CBDA and Δ^9 THCA standards, these compounds are analysed using respectively the CBD and Δ^9 THC standard response factors.

5

d) Test solutions

Diluted test solutions are made up in methanol and should contain analytes in the linear working range of 0.02-0.2 mg/ml.

10

e) Chromatography Acceptance Criteria:

The following acceptance criteria are applied to the results of each sequence as they have been found to result in adequate resolution of all analytes (including the two most closely eluting analytes CBD and CBDA)

15

Table 1- Retention time windows and Relative Retention Time (RRT) to Δ^9 THC for each analyte

20

Cannabinoid	Retention time (minutes)	RRT (THC)
CBD	5.1-5.8	0.58
CBN	7.4-8.3	0.83
Δ^9 THC	9.0-10.0	1.00
CBDA	5.5-6.2	0.615
Δ^9 THCV	5.9-6.2	0.645
CBC	11.6-12.8	1.30
Δ^9 THCA	14.6-16.0	1.605

25

30

Table 2- Peak Shape (Symmetry Factor according to British Pharmacopoeia method)

35

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Cannabinoid	Symmetry factor
CBD	<1.30
CBN	<1.25
Δ^9 THC	<1.35

5

f) Data Processing

10 Cannabinoids can be subdivided into neutral and acidic- the qualitative identification can be performed using the DAD dual wavelength mode. Acidic cannabinoids absorb strongly in the region of 220nm-310nm. Neutral cannabinoids only absorb strongly in the region of 220nm.

15

Routinely, only the data recorded at 220 nm is used for quantitative analysis.

20 The DAD can also be set up to take UV spectral scans of each peak, which can then be stored in a spectral library and used for identification purposes.

25 Data processing for quantitation utilises batch processing software on the Hewlett Packard Chemstation.

g) calculation:
Chromatographic purity of cannabinoid samples is calculated as a % of total cannabinoid content by area
30 normalization.

Capillary gas chromatography (GC) analysis

a) Chromatography equipment and conditions
35

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	Equipment	Agilent (HP) 5890 or 6890 GLC system with HP7673 Autosampler and FID detector
5	GLC column	SE54 (EC5) 30m x 0.32mm i.d. (Alltech) phase thickness 0.25 μ m
	Flow rate	Constant pressure (10.3 psi). Normal initial flow rate 34cm sec ⁻¹ (2.0 ml min ⁻¹)
10	Column oven	70°C initially then ramp 5°C min ⁻¹ to 250°C. Hold at 250°C for 15 minutes.
	Injector temp	250°C
	Detector temp	325°C
	Injection Vol	1 μ l, split ratio 2.5:1
	Run time	45 minutes
15	Fuel gases	Hydrogen 40 ml min ⁻¹ Air 450 ml min ⁻¹ Helium 45 ml min ⁻¹

20 b) Standard preparation

Stock standard solutions of CBD, CBN and Δ^9 THC in methanol at approximately 1mg ml⁻¹ are stored at -20°C. Diluted working standards (0.1 mg/ml for Δ^9 THC and CBD and 0.01 mg/ml for CBN) are prepared in methanol from the stock standards and stored at -20°C (maximum period of twelve months after initial preparation). Allow an aliquot pipetted into an autosampler vial to equilibrate to room temperature prior to use in a GC assay.

c) Sample preparation

35 Samples of final products, i.e. "pure" cannabidiol, are diluted in methanol prior to HPLC analysis. Optimal dilutions may be determined empirically.

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Cannabis plant material samples are prepared by taking 100mg chopped dried material and treating this with 5 or 10ml of Methanol/Chloroform (9:1 v/v). Extract the sample in an ultrasonic bath for 15 minutes and allow to stand in the dark for 18 hours.

d) Chromatography procedure

Standard solutions are used to provide quantitative and retention time data. These can be typically injected in triplicate prior to the injection of any sample solutions and then singularly at suitable intervals during the run, with a maximum of 10 test samples in between standards.

Table 3-Retention times

THCV	33.7-34.5 minutes
CBD	35.6-36.3 minutes
Δ^9 THC	37.2-38.1 minutes
CBN	38.5-39.1 minutes

TLC analysis

The qualitative composition of final products and starting materials may also be monitored by TLC.

TLC uses both retention time and characteristic spot colour to effectively identify the cannabinoid/cannabinoid acid components in a complex mixture. Methanolic solutions of the final products and starting material, plus standards, are prepared for TLC. An aliquot is spotted onto a TLC plate, alongside suitable reference samples (e.g. for at least Δ^9 THC and CBD). Following exposure to Fast Blue B reagent, THC and THCA present as pink spots,

- 39 -

while CBD and CBDA are orange in colour. Neutrals can be distinguished from the acids by comparison of the R_f value to that obtained for the standards. Identity is confirmed by comparison of R_f and colour of the sample spot, to that obtained for the appropriate standard.

A typical TLC protocol is as follows:

10 a) Materials and methods

Equipment:

Application device capable of delivering an accurately controlled volume of solution i.e 1 µl capillary pipette or micro litre syringe.

TLC development tank with lid

Hot air blower

20

Silica gel G TLC plates (SIL N-HR/UV254), 200 µm layer with fluorescent indicator on polyester support.

Dipping tank for visualisation reagent.

25

Mobile phase 80% petroleum ether 60:80/20% Diethyl ether.

30 Visualisation reagent 0.1% w/v aqueous Fast Blue B salt BN (Sigma Corp) (100mg in 100ml de-ionised water).
An optional method is to scan at UV 254 and 365 nm.

35 b) Sample preparation

i) Herbal raw material

- 40 -

Approximately 200mg of finely ground, dried cannabis is weighed into a 10ml volumetric flask. Make up to volume using methanol:chloroform (9:1) extraction solvent.

5

Extract by ultrasound for 15 minutes. Decant supernatant and use directly for chromatography.

ii) Final products

10

The final products (crystalline CBD) are dissolved in methanol to a suitable concentration (which may be determined empirically) then used directly for chromatography. All sample preparations should produce a final concentration of about 0.5 mg/ml.

15

iii) Botanical drug substance

20

Accurately weigh approximately 50 mg of botanical drug substance into a 25 ml volumetric flask. Dissolve to make volume with HPLC grade methanol.

c) Standards

25

0.1 mg/ml Δ^9 -THC in methanol (Sigma).
0.1mg/ml CBD in methanol (Sigma).

30

The standard solutions are stored frozen at -20°C between uses and are used for up to 12 months after initial preparation.

d) Test solutions and method

35

Apply to points separated by a minimum of 10mm.

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- i) either 5 μ l of herb extract or 1 μ l of pure cannabinoid/enriched extract solution or 1 μ l of diluted column eluate as appropriate,
- ii) 5 μ l of 0.1 mg/ml Δ^9 -THC in methanol standard solution,
- 5 iii) 5 μ l of 0.1mg/ml CBD in methanol standard solution.

Dry the prepared plate with a hot air blower.

10

Place the base of the TLC plate in a development tank containing the mobile phase and saturated with vapour.

15

Elute the TLC plate through a distance of 8cm, then remove the plate. Allow solvent to evaporate from the plate and then repeat the elution for a second time (double development). Remove plate and allow it to dry in air.

20

The entire plate is briefly immersed in the Fast Blue B reagent until the characteristic red/orange colour of cannabinoids begins to develop. The plate is removed and allowed to dry under ambient conditions in the dark.

25

Cannabinoids will give an orange-purple colour:

- | | | |
|-----------------------------------|------|--------------------------|
| Cannabidiol | CBD | orange (fastest running) |
| Δ^9 Tetrahydrocannabinol | THC | pink |
| Cannabinol | CBN | purple |
| 30 Cannabichromene | CBC | pink purple |
| Cannabigerol | CBG | orange |
| Δ^9 tetrahydrocannabivarin | THCV | purple |

35

The corresponding acids form streaks of the same colour as the neutral component spots. The acids run at lower R_f .

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Claims:

1. A cannabinoid acid for use as an active pharmaceutical substance.
2. Use of a cannabinoid acid for the manufacture of a medicament for the treatment of nausea, vomiting, emesis, motion sickness or like conditions.
3. A cannabinoid acid for use in treating nausea, vomiting, emesis, motion sickness or like conditions.
4. CBDA or CBDVA for use as an active pharmaceutical substance.
5. Use of CBDA or CBDVA for the manufacture of a medicament for the treatment of nausea, vomiting, emesis, motion sickness or like conditions.
6. A method of treating nausea, vomiting, emesis, motion sickness or like conditions in a mammal comprising administering a pharmaceutically effective amount of or an extract containing a pharmaceutically effective amount of CBDA or CBDVA.
7. CBDA or CBDVA for use in treating nausea, vomiting, emesis, motion sickness or like conditions.
8. Use of a cannabis extract rich in CBD and/or CBDA and/or the propyl variants CBDV and/or CBDVA, for the manufacture of a medicament for the treatment of nausea, vomiting, emesis, motion sickness or like conditions.
9. A method of treating nausea, vomiting, emesis, motion sickness or like conditions in a mammal comprising administering a pharmaceutically effective

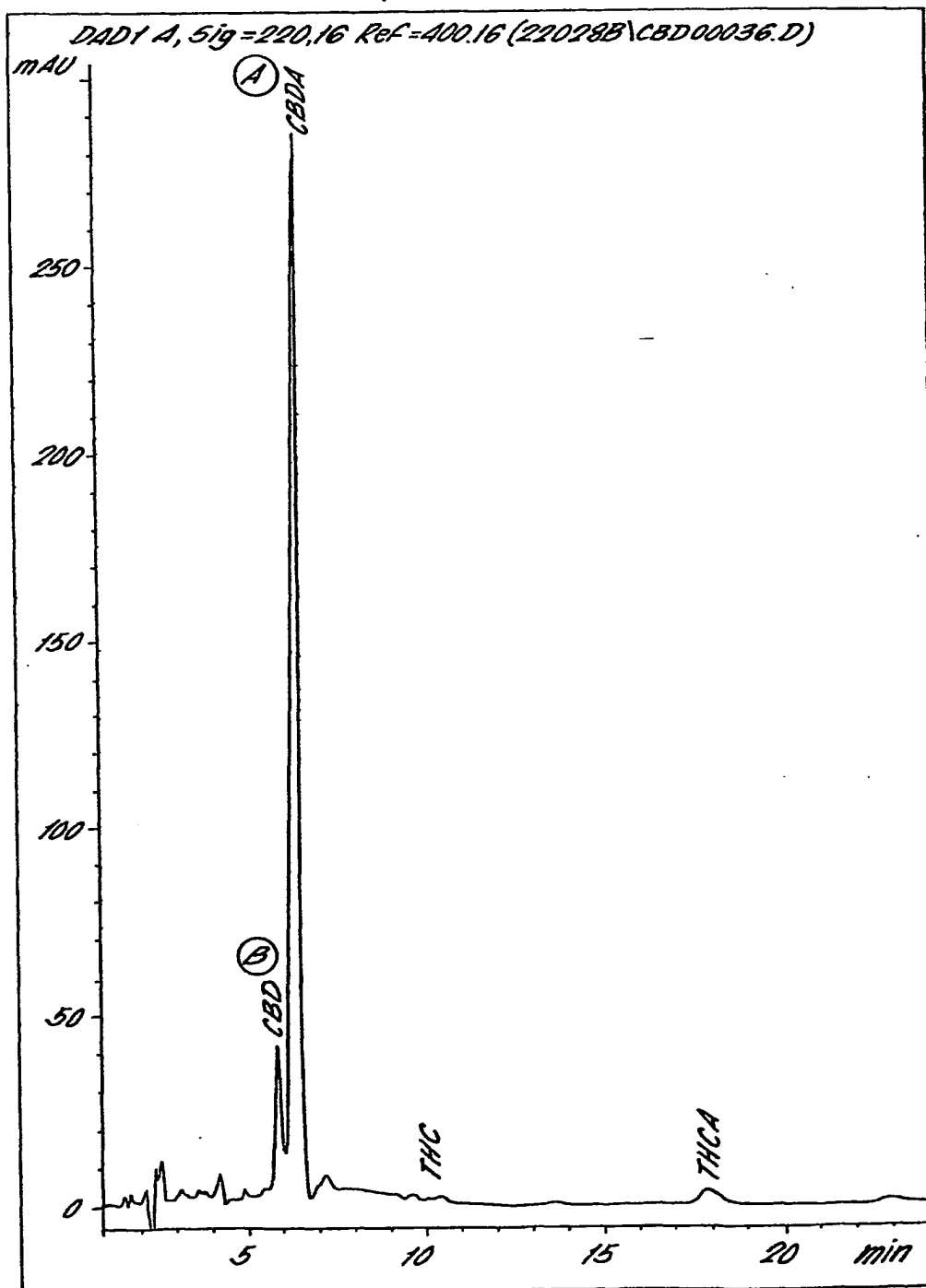
- 43 -

amount of, or an extract containing a pharmaceutically effective amount of CBD and/or CBDA and/or the propyl variants CBDV and/or CBDVA.

- 5 10. The use of claim 8 or claim 9 wherein the extract comprises greater than 2% w/w of CBD and/or CBDA and/or the propyl variants CBDV and/or CBDVA, more particularly greater than 5%, more preferably still greater than 7%.
- 10 11. A cannabis extract rich in CBD and/or CBDA and/or the propyl variants CBDV and/or CBDVA for use in treating nausea, vomiting, emesis, motion sickness or like conditions.
- 15 12. A pharmaceutical composition comprising CBDA and/or CBDVA and a pharmaceutically acceptable carrier or diluent.
- 20 13. Use of CBD or CBDV for the manufacture of a medicament for the treatment of nausea, vomiting, emesis, motion sickness or like conditions.
- 25 14. Use of CBD or CBDV as claimed in claim 13 wherein the CBD or CBDV is present with other cannabinoids derived as a plant extract.
- 30 15. Use of CBD or CBDV as claimed in claim 14 wherein the other cannabinoids include THC or THCV.
- 35 16. Use of CBD or CBDV as claimed in claim 13 wherein the CBD or CBDV is substantially pure.
17. Use of CBD or CBDV as claimed in claim 13 wherein the CBD or CBDV is synthetic.
18. CBD or CBDV for use in treating nausea, vomiting, emesis, motion sickness or like conditions.

*FIG. 1 HPLC CHROMATOGRAM OF AC 1494
METHANOLIC EXTRACT OF G5 HERB*

CURRENT CHROMATOGRAM(S)



SUBSTITUTE SHEET (RULE 26)

FIG. 2. HPLC CHROMATOGRAM OF AC 1495
METHANOLIC EXTRACT OF G2 HERB

CURRENT CHROMATOGRAM(5)

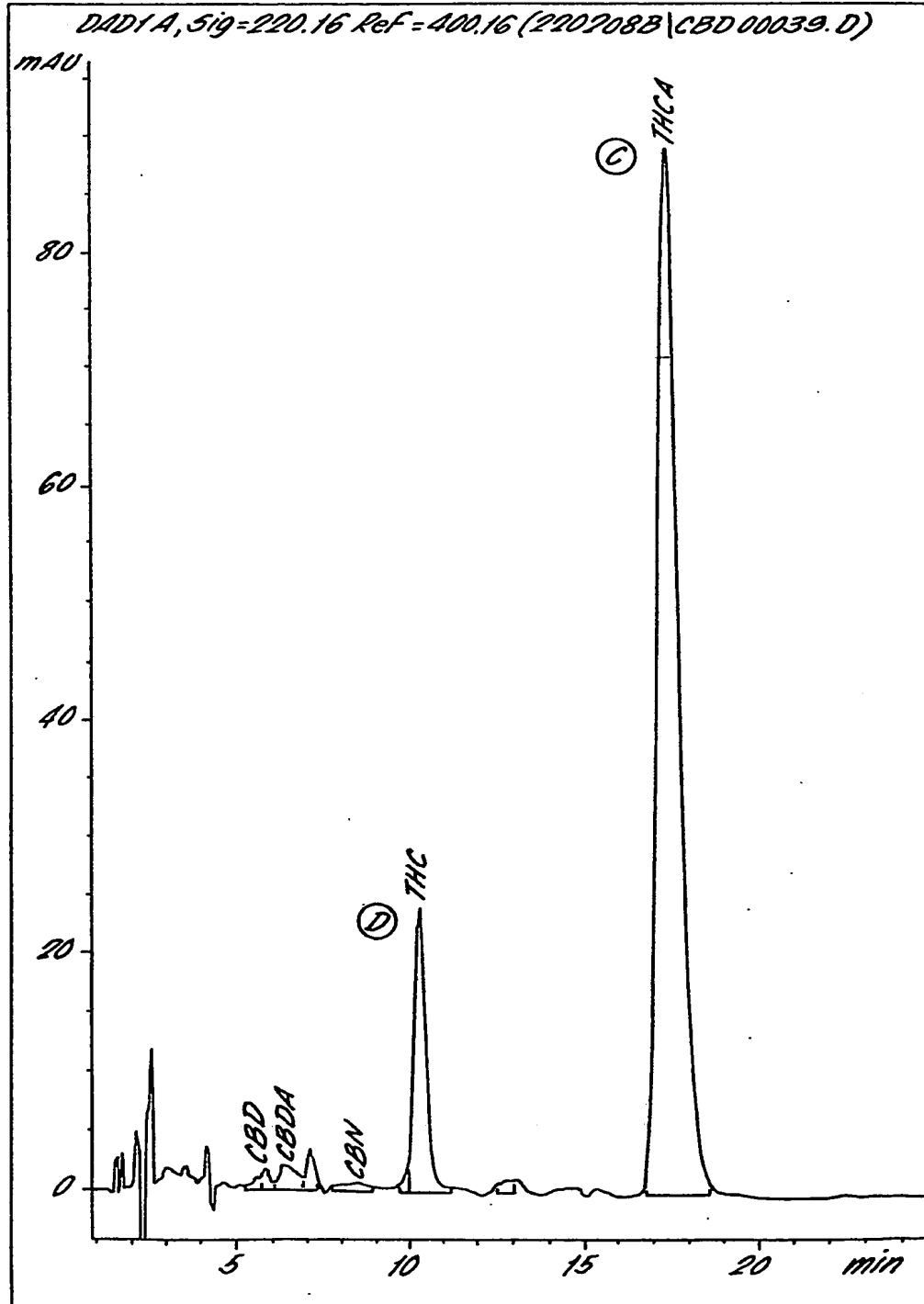


FIG. 3.

TLC COMPARISON OF METHANOLIC EXTRACTS OF
G2 HERB (THC) AND G5 HERB (CBD) TO LC02
EXTRACTS OF CORRESPONDING CHEMOVAR
i.e. BDS(THC) AND BDS(CBD)

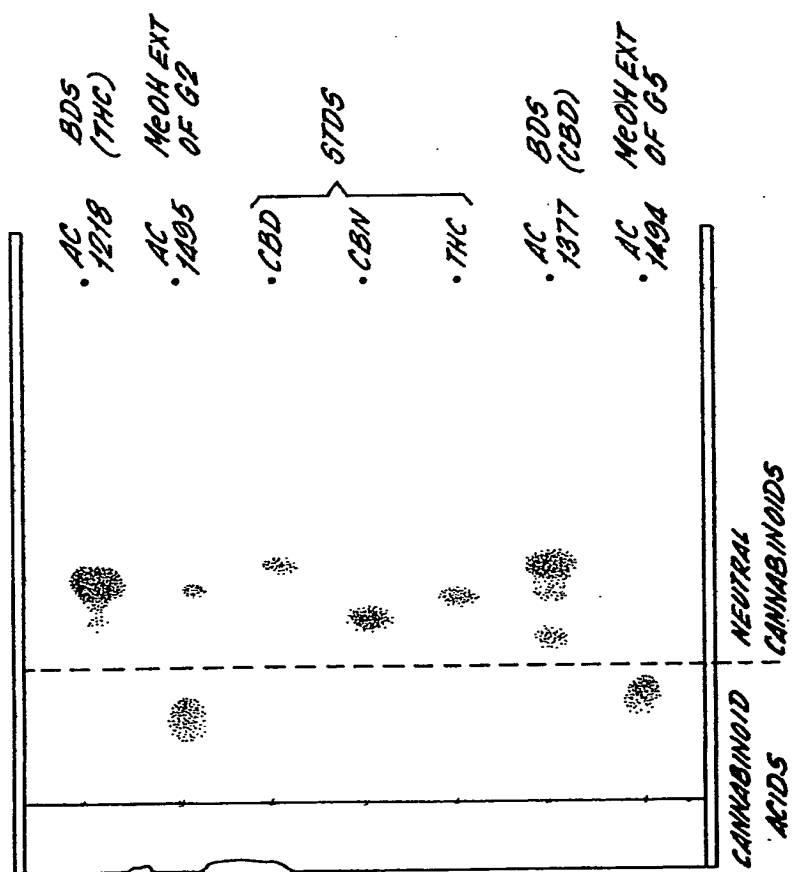


FIG. 4.

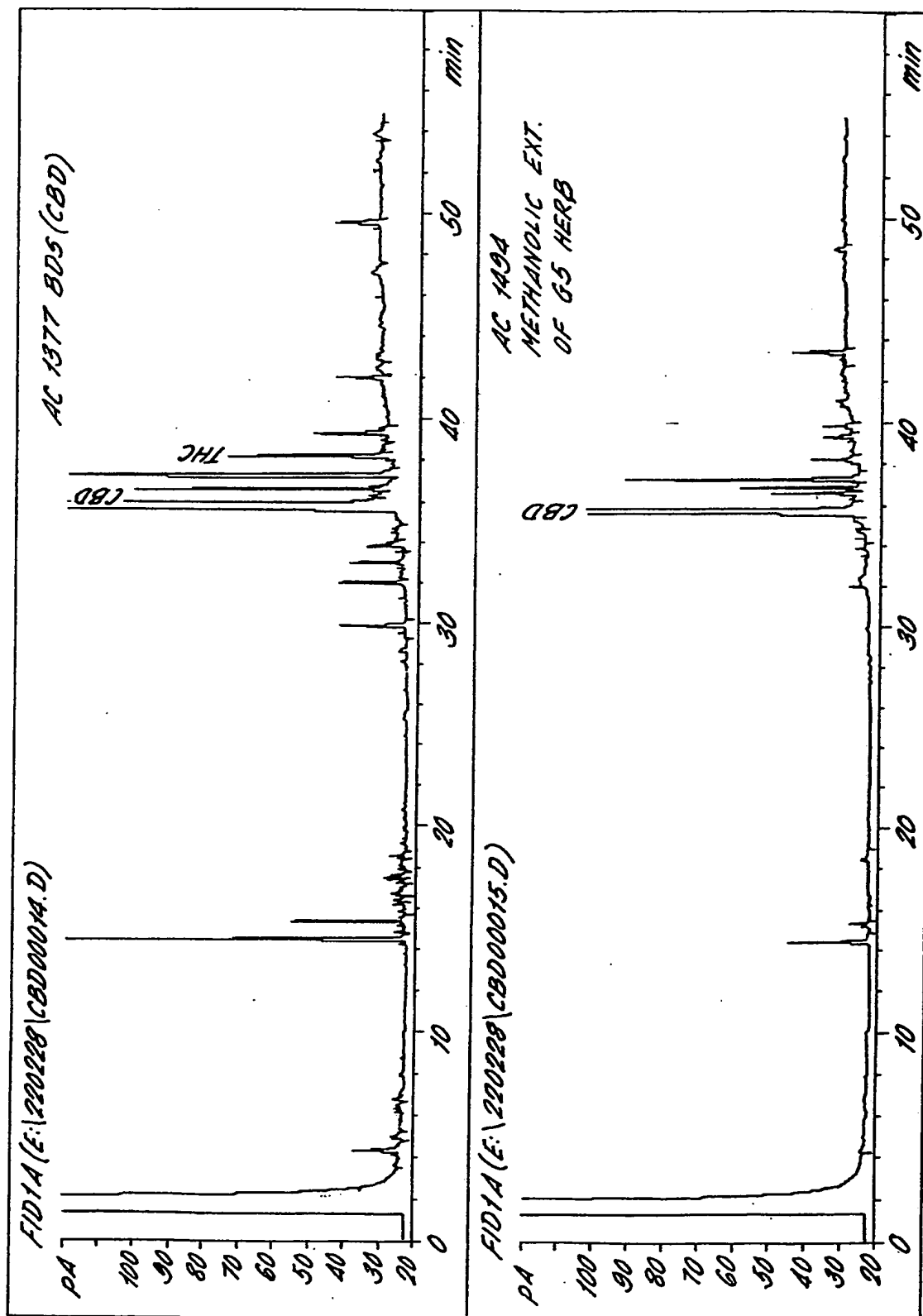
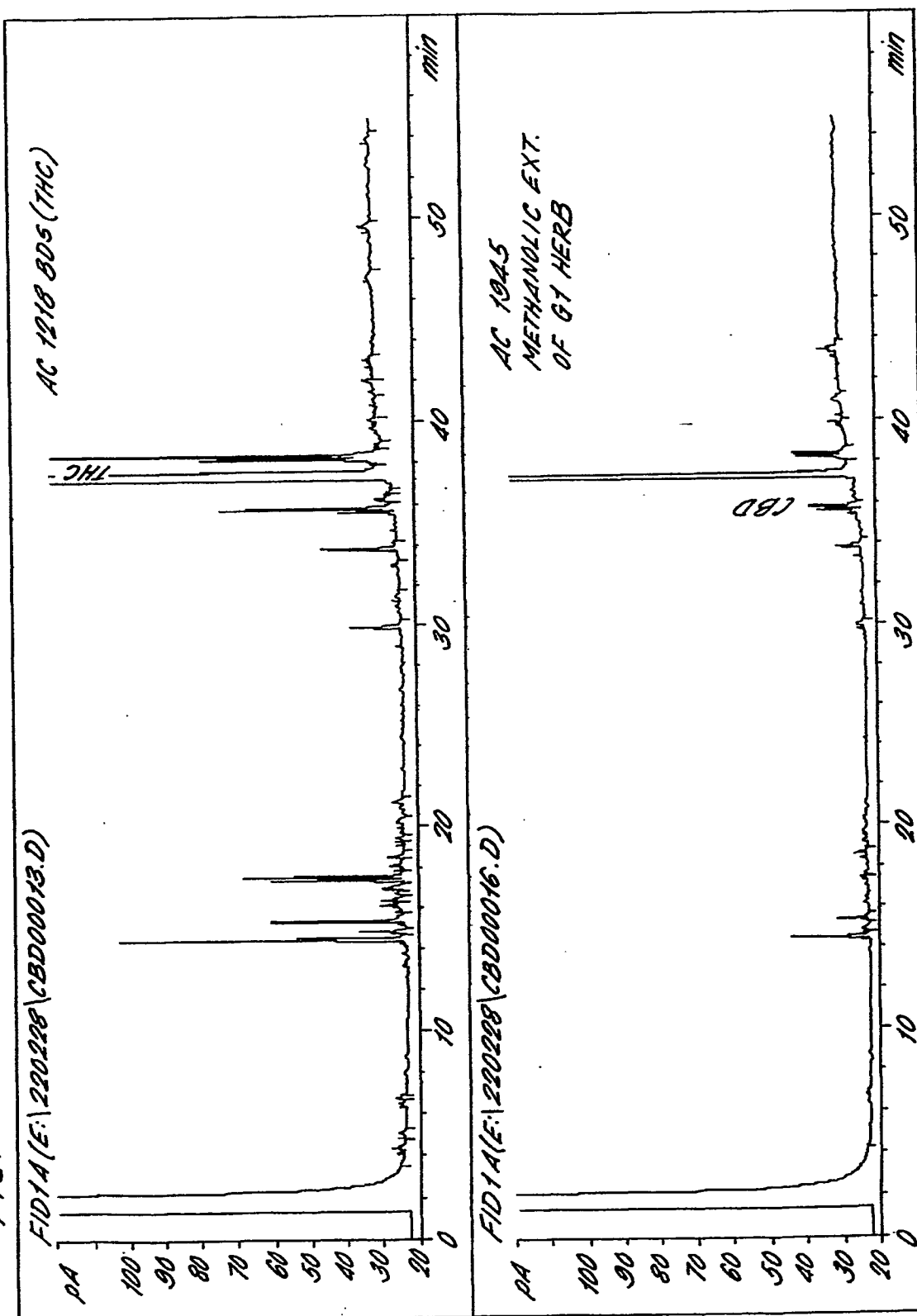


FIG. 5.



SUBSTITUTE SHEET (RULE 26)

INTERNATIONAL SEARCH REPORT

 Internat. Application No
 PCT/LU 3/00451

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 A61K31/05 A61K31/192 A61K35/78 A61P1/08

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K A61P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, EMBASE, BIOSIS, MEDLINE, PASCAL, SCISEARCH, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 4 847 290 A (BURSTEIN SUMNER) 11 July 1989 (1989-07-11) column 1, line 45-65 column 3, line 40-47	1-3
X	WO 93 05031 A (YISSUM RES DEV CO) 18 March 1993 (1993-03-18)	1-3
Y	claims 16-19	1-18
X	US 5 434 295 A (MECHOULAM RAPHAEL ET AL) 18 July 1995 (1995-07-18) column 7, line 34-49; claim 20; example 6	1-3
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☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
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- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
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Date of the actual completion of the international search

14 May 2003

Date of mailing of the international search report

06/06/2003

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Friederich, M

INTERNATIONAL SEARCH REPORT

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 PCT/1... 3/00451

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X	WO 01 95899 A (MECHOULAM RAPHAEL ;YISSUM RES DEV CO (IL); FRIDE ESTER (IL); HANUS) 20 December 2001 (2001-12-20) page 2, paragraph 3 -page 3, paragraph 1	1,3
X	YOTORIYAMA M ET AL: "COMPARISON OF PHARMACOLOGICAL ACTIVITY IN MICE OF DIFFERENT CANNABIS EXTRACTS FROM CBDA AND THCA STRAINS" EISEI KAGAKU, vol. 37, no. 6, 1991, pages 507-511, XP009010606 ISSN: 0013-273X * the whole document *	4,7,12
X	MOLNAR J ET AL: "Membrane associated antitumor effects of crocine-, ginsenoside- and cannabinoid derivates." ANTICANCER RESEARCH, vol. 20, no. 2A, March 2000 (2000-03), pages 861-868, XP009010607 ISSN: 0250-7005 figure 2; table 6	4,7,12
P,X	PARKER LINDA A ET AL: "Cannabidiol, a non-psychoactive component of cannabis and its synthetic dimethylheptyl homolog suppress nausea in an experimental model with rats." NEUROREPORT. ENGLAND 16 APR 2002, vol. 13, no. 5, 16 April 2002 (2002-04-16), pages 567-570, XP009010573 ISSN: 0959-4965 * the whole document *	13,16-18
P,X	WO 02 064109 A (GW PHARMA LTD ;WHITTLE BRIAN (GB); GUY GEOFFREY (GB)) 22 August 2002 (2002-08-22) examples	13-18

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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X	WO 99 52524 A (FELDMANN MARC ;MALFAIT ANNE MARIE (GB); MATHILDA & TERENCE KENNEDY) 21 October 1999 (1999-10-21) page 5, line 16,17; examples page 6, line 9-11 ---	18
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Y	YAMAMOTO I ET AL: "Recent advances in the metabolism of cannabinoids" INTERNATIONAL JOURNAL OF BIOCHEMISTRY AND CELL BIOLOGY 1995 UNITED KINGDOM, vol. 27, no. 8, 1995, pages 741-746, XP002241104 ISSN: 1357-2725 page 742, column 2, paragraph 1 page 744, column 2, paragraph 2 ---	1-18
A	ADAMS R ET AL: "Structure of Cannabidiol. XII. Isomerization to Tetrahydrocannabinols" JOURNAL OF THE AMERICAN CHEMICAL SOCIETY, AMERICAN CHEMICAL SOCIETY, WASHINGTON, DC, US, vol. 63, 1941, pages 2209-2213, XP002208081 ISSN: 0002-7863 -----	

INTERNATIONAL SEARCH REPORT

Int. Application No.
B 03/00451

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: —
because they relate to subject matter not required to be searched by this Authority, namely:
see FURTHER INFORMATION sheet PCT/ISA/210
2. ☐ Claims Nos.: —
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.: —
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

Continuation of Box I.1

Although claims 6 and 9 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

Continuation of Box I.1

Rule 39.1(iv) PCT - Method for treatment of the human or animal body by therapy

INTERNATIONAL SEARCH REPORT

Internat
PCT,Application No
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